Determination of neonicotinoid insecticide residues in bees, pollen and nectar with LC-MS/MS

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Seed coating with a neonicotinoid insecticide negatively affects wild bees

Nature (2015) 521: 77-80

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Background

- Spring sown oilseed rape (just over 40 % in Sweden)
- Neonicotinoid (mostly clothianidin) seed dressing to protect seedlings
- Lack of well designed replicated field studies, particularly for non-honeybees

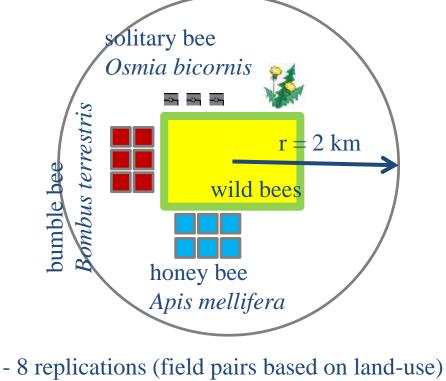


Flea beetle (*Phyllotreta* sp.) Photo: Maj Rundlöf

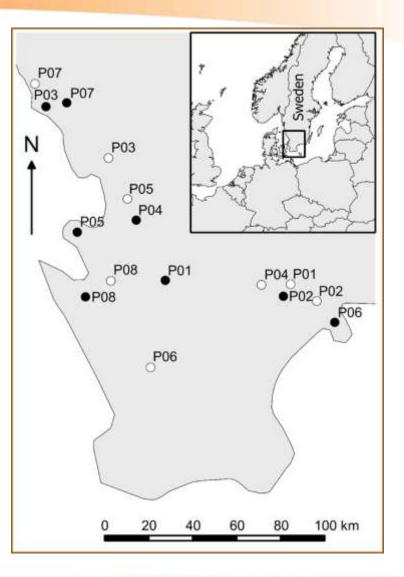


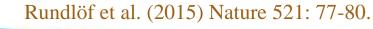


Landscape ecotoxicology experiment



- random assignment to treatment/control
- treatment blinded during field work



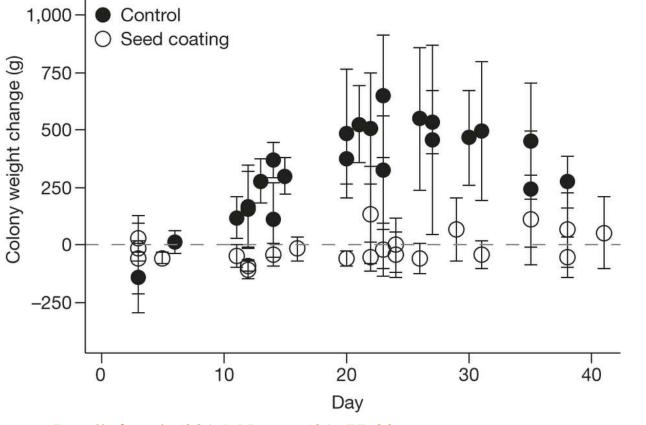


SLU



Results

The clothianidin treatment was negatively related to *Bombus terrestris* colony growth





Photos: Maj Rundlöf

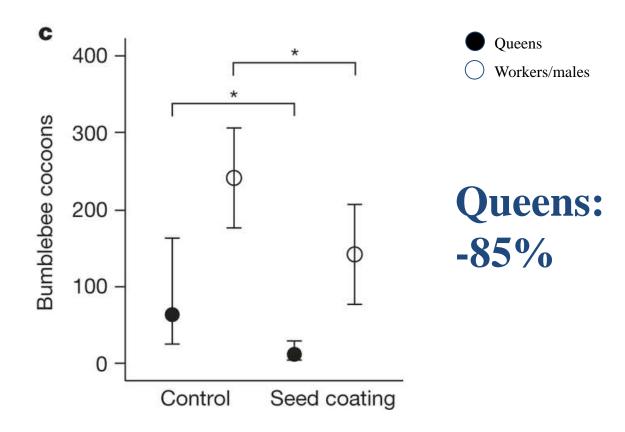


Rundlöf et al. (2015) Nature 521: 77-80.





...and Bombus terrestris reproduction



Photos: Maj Rundlöf



Rundlöf et al. (2015) Nature 521: 77-80.

SLU



Reduced nesting of solitary bee Osmia bicornis (red mason bee) in tubes placed close to the fields

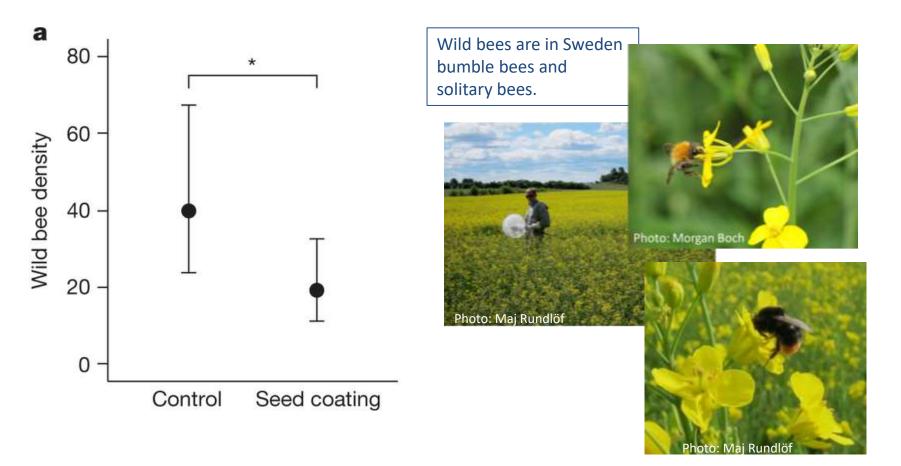


Rundlöf et al. (2015) Nature 521: 77-80.





Reduced wild bee density in treated fields (transects)

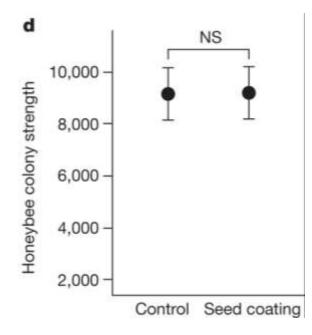


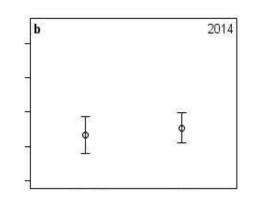
Rundlöf et al. (2015) Nature 521: 77-80.





The clothianidin seed treatment had no significant influence on *Apis mellifera* colony strength









Not always possible to extrapolate ecotoxicology results for honey bees to wild bees or other insect species.

Rundlöf et al. (2015) Nature 521: 77-80; Rundlöf et al. (2015) Project report, Lund university





Chemical analysis



5 neonicotinoid insecticides studied:

Acetamiprid Imidacloprid Clothianidin Thiacloprid Thiamethoxam

Internal standards: Imidacloprid D4 Clothianidin D3 Isoproturone D6 (used for acetamiprid, thiacloprid and thiamethoxam)

Agilent 6460 LC-MS/MS, on-line SPE (C18 and polymer) C18 analytical column, methanol gradient in ammonium formate [1]



¹C. Jansson and J. Kreuger 2010 J. AOAC Int., Volume 93, Number 6, 1732-1747



Sample preparation bees (and pollen)

- 24 bees weighed and homogenized with drying agent, subsample corresponding to four bees
- internal standard to subsample homogenate
- extract twice under strong sonication with
 7:3 mixture of acetone and ethyl acetate
- dispersive solid phase extraction, C18 and PSA
- evaporate to dryness at 40°C under nitrogen flow
- residue dissolved in 150 μl acetonitrile
- 10 µl injected on LC-MS/MS

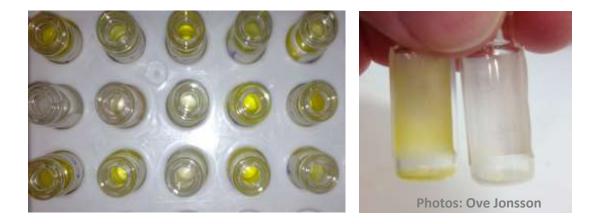


Vibra-Cell VCX 130 (Sonics)





Bees and pollen are varying matrices



Extracts from different bee samples.

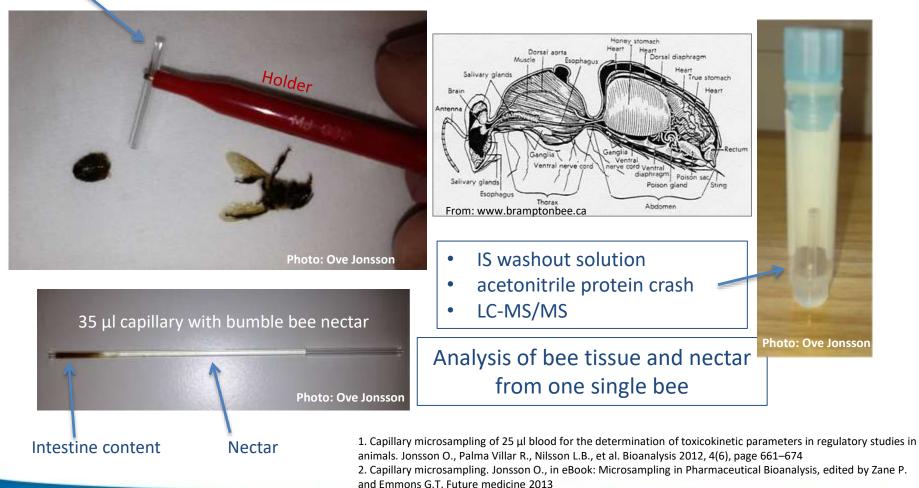




Collection and handling of nectar samples

Capillary with exact 8 µl nectar (deviation and %RSD <1%)

8 μl capillary in 1 ml tube Capillary Microsampling (CMS) [1,2]





Clothianidin field exposure

(ng/g or ml) 2013	Control (C)		Seed coating (T)		N (C/T)
	range	mean ± se	range	mean ± se	
Honey bees	<lod-0,89< td=""><td>0,13 ± 0,11</td><td>0,35-4,9</td><td>$2,4 \pm 0,50$</td><td>8/8</td></lod-0,89<>	0,13 ± 0,11	0,35-4,9	$2,4 \pm 0,50$	8/8
Pollen from honey bees	<lod< td=""><td><lod< td=""><td>6,6-23</td><td>14 ± 1,8</td><td>6/8</td></lod<></td></lod<>	<lod< td=""><td>6,6-23</td><td>14 ± 1,8</td><td>6/8</td></lod<>	6,6-23	14 ± 1,8	6/8
Nectar from honey bees	<lod-0,61< td=""><td>0,11 ± 0,080</td><td>6,7-16</td><td>10 ± 1,3</td><td>8/8</td></lod-0,61<>	0,11 ± 0,080	6,7-16	10 ± 1,3	8/8
2014					
Honey bees	<lod< td=""><td><lod< td=""><td>0,15-1,5</td><td>$\textbf{1,1} \pm \textbf{0,20}$</td><td>4/6</td></lod<></td></lod<>	<lod< td=""><td>0,15-1,5</td><td>$\textbf{1,1} \pm \textbf{0,20}$</td><td>4/6</td></lod<>	0,15-1,5	$\textbf{1,1} \pm \textbf{0,20}$	4/6
Pollen from honey bees	<lod< td=""><td><lod< td=""><td>2,4-16</td><td>$\textbf{6,1} \pm \textbf{2,0}$</td><td>4/6</td></lod<></td></lod<>	<lod< td=""><td>2,4-16</td><td>$\textbf{6,1} \pm \textbf{2,0}$</td><td>4/6</td></lod<>	2,4-16	$\textbf{6,1} \pm \textbf{2,0}$	4/6
Nectar from honey bees	<lod< td=""><td><lod< td=""><td>2,6-9,8</td><td>4,9 ± 1,1</td><td>4/6</td></lod<></td></lod<>	<lod< td=""><td>2,6-9,8</td><td>4,9 ± 1,1</td><td>4/6</td></lod<>	2,6-9,8	4,9 ± 1,1	4/6

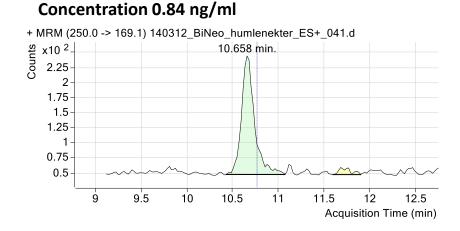
LOD: 0.080-0.50, LOQ: 0.25-1.5 ng/g or ng/ml

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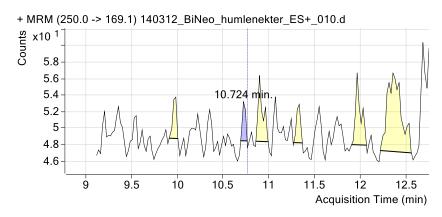




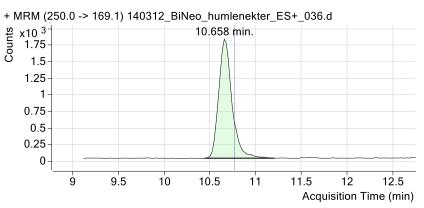
Clothianidin chromatogram from nectar samples



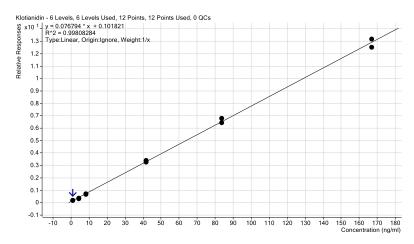
Nectar sample from control field



Highest concentration in nectar sample 20 ng/ml



Calibration curve clothianidin in nectar



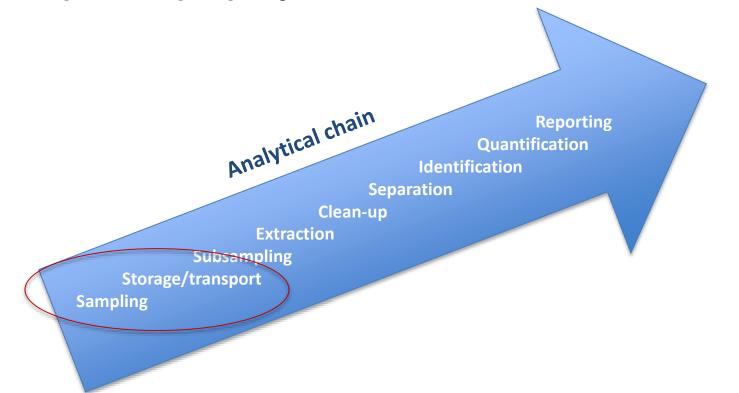




1. Individual exposure variation within a bee society Representative sampling

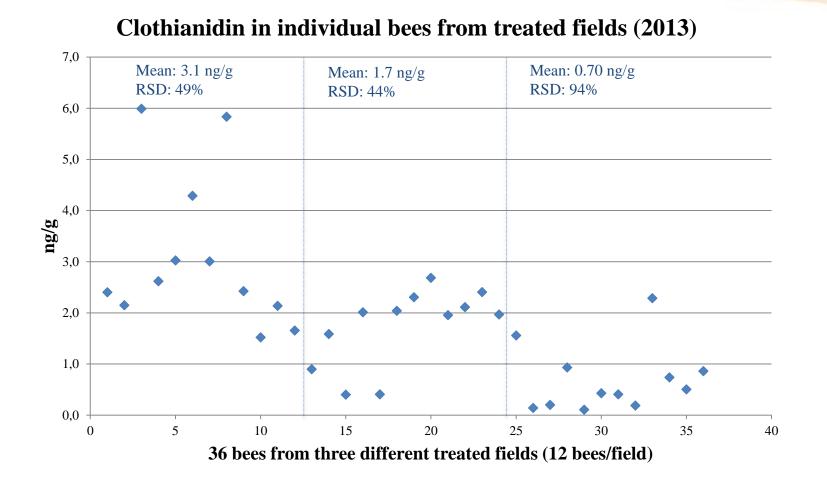
2. Stability of pesticides in stored bees

Important step in quality assurance







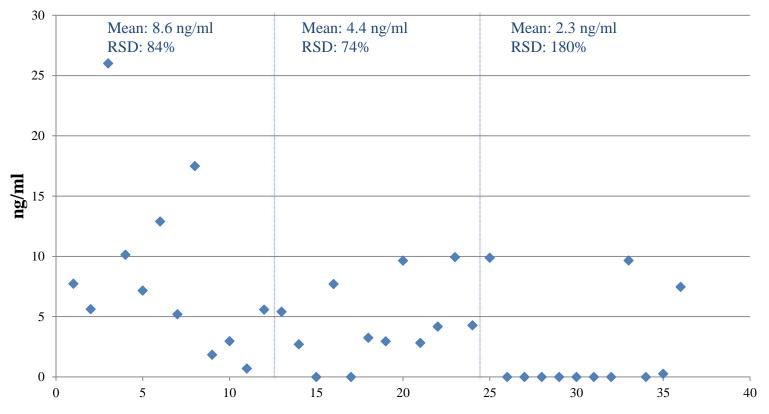


Overall mean: 1.8 ng/g RSD: 77%





Clothianidin in nectar from individual bees, collected in three treated fields (2013)



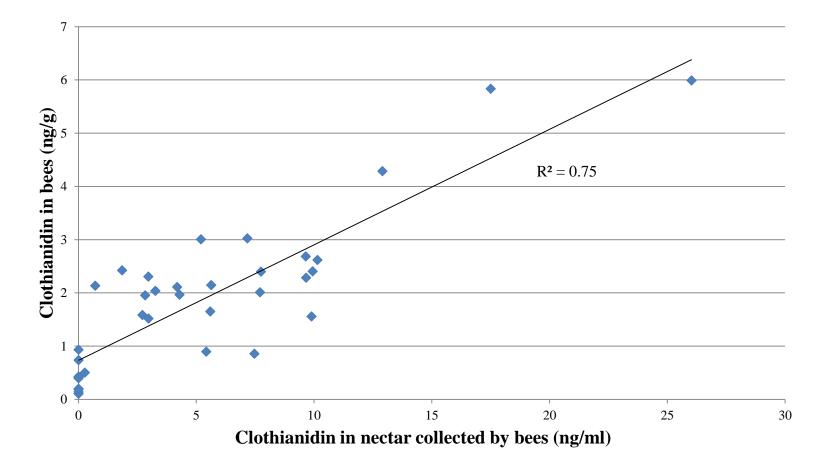
Nectar from 36 bees from three treated fields, (12 from each field)

Overall mean: 5.1 ng/ml RSD: 111%



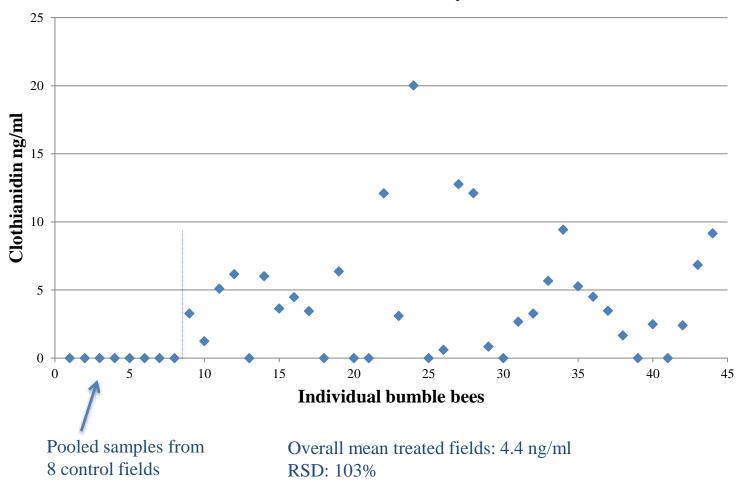


Clothianidin concentrations in 36 bees and the nectar they collected from treated fields









Clothianidin in nectar collected by bumble bees





Validation data

Field study

Bee and pollen samples:

Calibration curve in solvent and matrix spiking experiments at two concentrations.

RSD bees 0.9-18% and pollen 2.4-19%,

for both matrices, concentration levels and all five neonicotinoids.

(clothianidin 4.7% and 1.0% for low and high spiking level in bees and 7.3% and 12% in pollen).

Storage stability experiment

Bees:

Mean RSD for spiked homogenate 3-8%

For all five Neonicotinoids in five batches over 21 months. (n=3 per batch, one concentration level)

Nectar samples:

Calibration curve and Quality Controls (QC) at two levels in nectar collected by bees (matrix match)

QC Accuracies of 89-101% and QC RSD of 1.1-8.2%

for both concentration levels and all five neonicotinoids.





Pesticide stability in bees after individual feeding

Individual feeding with 10 μl sugar solution containing pesticide mix – Relevant spiking! Evaluate stability *in vivo*, in room temperature (dead bees) and after long time freeze storage at -20°C



Professor Ingemar Fries, SLU, feeds a bee.

Freezer and room temp: All five neonicotinoids were stable in freezer for 21 months and at room temperature for 22 hours.

In vivo:

Clothianidin was stable for 1 h while acetamiprid, imidacloprid, thiacloprid and thiamethoxam showed degradation with 44, 44, 43 and 22%, respectively.

Each estimate was based on triplicate samples and four bees per sample (in total 12 bees)





Acknowledgement pesticides in bee matrices

Swedish EPA and the Centre for Chemical Pesticides for financial support to develop analytical methods

Maj Rundlöf for support with field study slides

Prof. Ingemar Fries, SLU, for feeding bees



Photo: Maj Rundlöf





Micro sampling side track



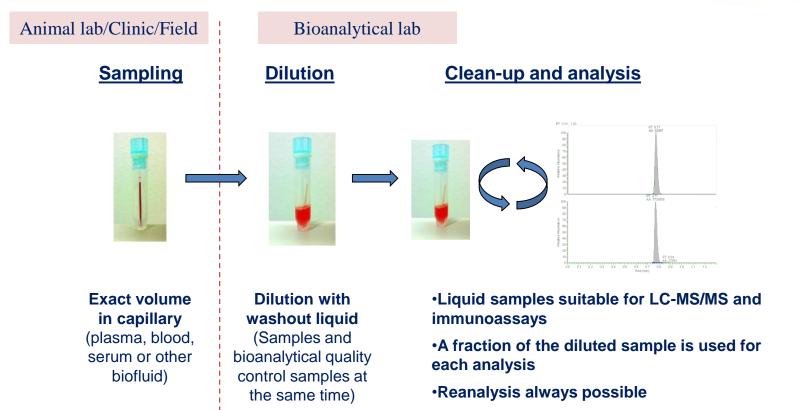




Capillary MicroSampling (CMS) of blood, plasma and serum

- Exposure of drug candidate and/or biomarker determinations
- Repeated sampling from small rodents including juveniles
- Compatible with LC-MS and immunoassays

Basic CMS principle

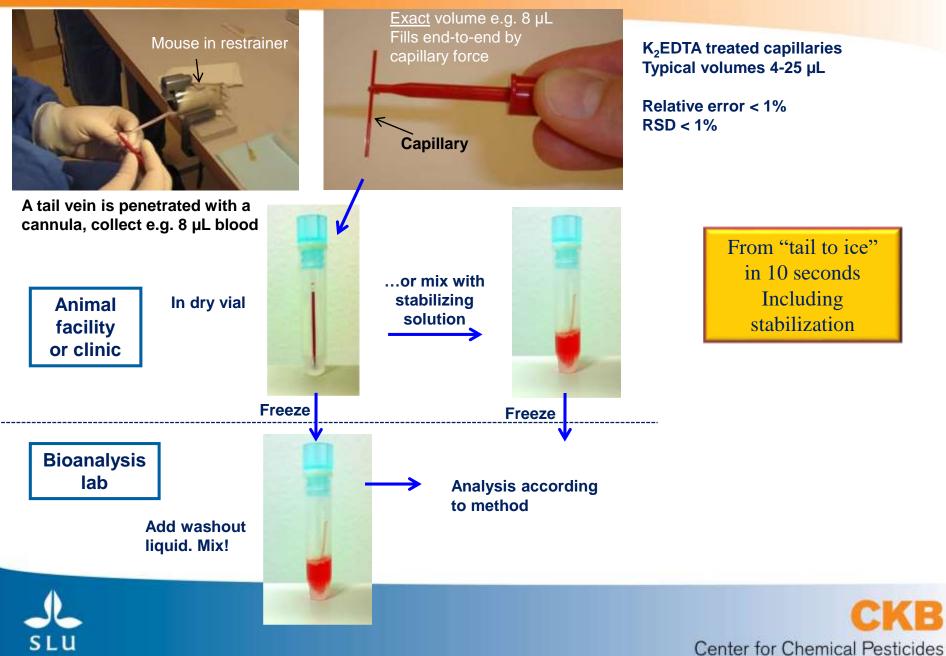


•Standard lab equipment for liquid samples





CMS of blood

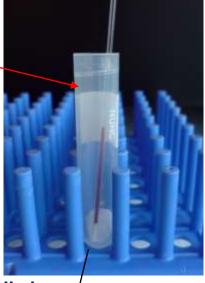


CMS of plasma (serum)

~32 µL blood in K₂EDTA haematocrit tube (plain glass for serum sampling)

Plug with wax

Centrifugation 1500 g for 10 min



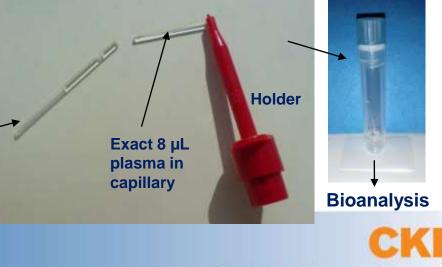
Agilent Technologian

Cut above the blood cell phase using a capillary cutter.

Erythrocytes Leucocytes & thrombocytes Wax plug

Typical volumes 8 μL (4 μL backup) plasma from ~32 μL blood

An exact volume of plasma is collected with a capillary from the end of the haematocrit tube. Put capillary in tube or plate



Center for Chemical Pesticides

Can pesticides affect breeding success in agricultural bird species? Collaboration: SLU (OMK and Ecology dep.) + Kvismare bird observatory













Pesticides in blood?





Sampling site on birds

Here male redstart (rödstjärt)



Approximately 100 micro samples successfully collected in the field









Determination of pesticides in blood: Capillary microsampling, protein precipitation and LC-MS/MS

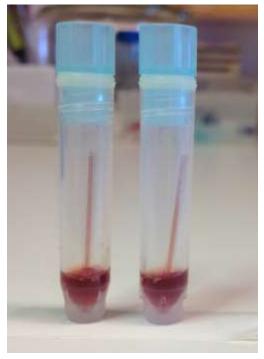
8 µl blood in K₂EDTA capillary 16 µl 25% ACN 56 µl IS in ACN Approx. 65 µl Mix and spin! Transferred to LC-injection vial



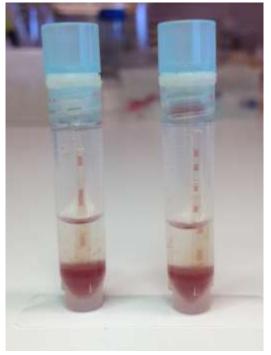


Liquid-liquid extraction of pesticides in blood

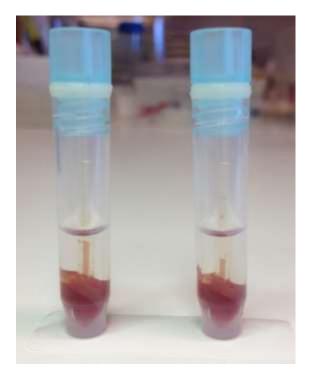
Blood (8 µl) mixed with internal standard (IS) in water (80 µl)



Organic solvent* (250 µl) added and mixed for liquid/liquid extraction



After centrifugation 2000 x g for 2 minutes

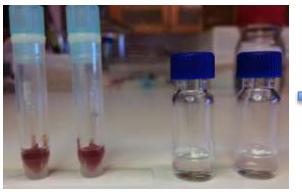


*In this case cyclohexane:ethylacetate 1:1 (v:v)

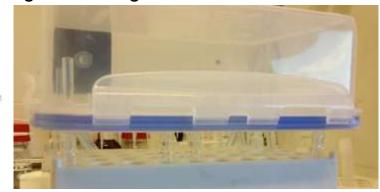




Organic solvent extracts transferred to new vials



Extracts evaporated under gentle nitrogen flow

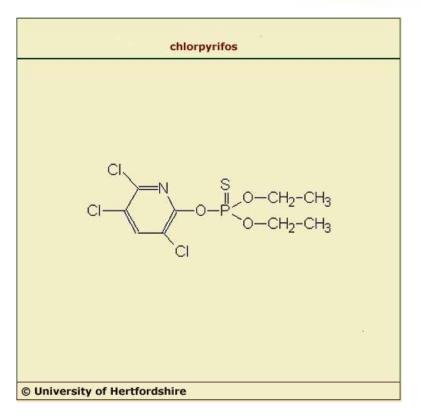




Final extracts reconstituted in 80 µl cyclohexane:acetone 9:1 ready for GC-MS injection







Mode of action Non-systemic with contact and stomach action. Acetylcholinesterase (AChE) inhibitor.





Thank you for your time!

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